

Toward the synthesis of fine chemicals from lactose: preparation of *D*-xylo and *L*-lyxo-aldohexos-5-ulose derivatives[‡]

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Abstract – The transformation of (5*R*)-2,6-di-*O*-benzyl-5-*C*-methoxy-β-*D*-galactopyranosyl-(1→4)-2,3:5,6-di-*O*-isopropylidene-*aldehydo-D*-glucose dimethyl acetal (**8**) into partially protected derivatives of *D*-xylo- and *L*-lyxo-aldohexos-5-ulose has been reported, applying appropriate epimerization methods to its 3'-*O*- and 4'-*O*-protected alcoholic derivatives.

Keywords: Lactose, *D*-xylo-Aldohexos-5-ulose, *L*-lyxo-Aldohexos-5-ulose, 1,5-Bis-glycopyranosides, Epimerization

1. Introduction

Lactose is the most abundant natural reducing disaccharide, obtained from whey, a by-product of the agro-industrial cheese production. Although its large worldwide availability, estimated in about 500,000 tons/year,^{2a} only a low percentage of recovered lactose is utilized, mainly in the food, feed and pharmaceutical fields. The chemical valorisation of lactose is achieved through simple transformations into commercially available products as lactobionic acid,² a component of the preservative solution for transplanting organs, lactitol,² a suitable component of sugar-free, reduced calories and low glycaemic products, and lactulose and galacto-oligosaccharides (GOS),² largely used in probiotic therapy.

Since lactose is cheap and there is a potential environmental risk connected with the uncontrolled dispersion of whey in freshwater, new synthetic channels are investigated in order to synthesise fine chemicals starting from this renewable raw material.

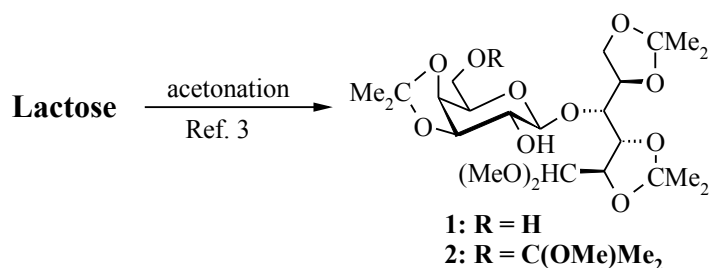


Figure 1. Polyacetone derivatives directly obtained by acetonation of lactose

[‡]Part 26 of the series “Chemical Valorisation of Milk-derived Carbohydrates”. For part 25 see Ref. 1.

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Recently, we have planned a synthetic strategy to elaborate the not reducing unit and not to modify the reducing one, as commonly done. This useful approach takes advantage of the large availability³ of the two polyacetonides **1** and **2** (Figure 1), which could be considered as simple β -D-galactopyranosides, due to the complete protection of the D-glucose unit. Aldohexos-5-uloses (**3**) represent an interesting, although yet poorly investigated class of dicarbonyl hexoses,⁴ useful synthetic intermediates for the preparation of high added value compounds such as azasugars⁵ and cyclitols, as inositols,⁶ or polyhydroxycyclopentanes.⁷

A general approach (Chart 1) to aldohexos-5-uloses (**3**) was developed⁸ using as key reaction the epoxidation-methanolysis of hex-4-enopyranosides of type **5**, in turn obtained from 3,4-O-isopropylidene- β -D-galactopyranosides (**6**).

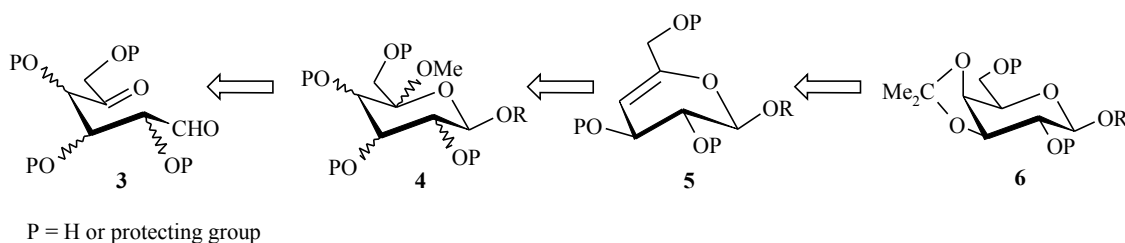
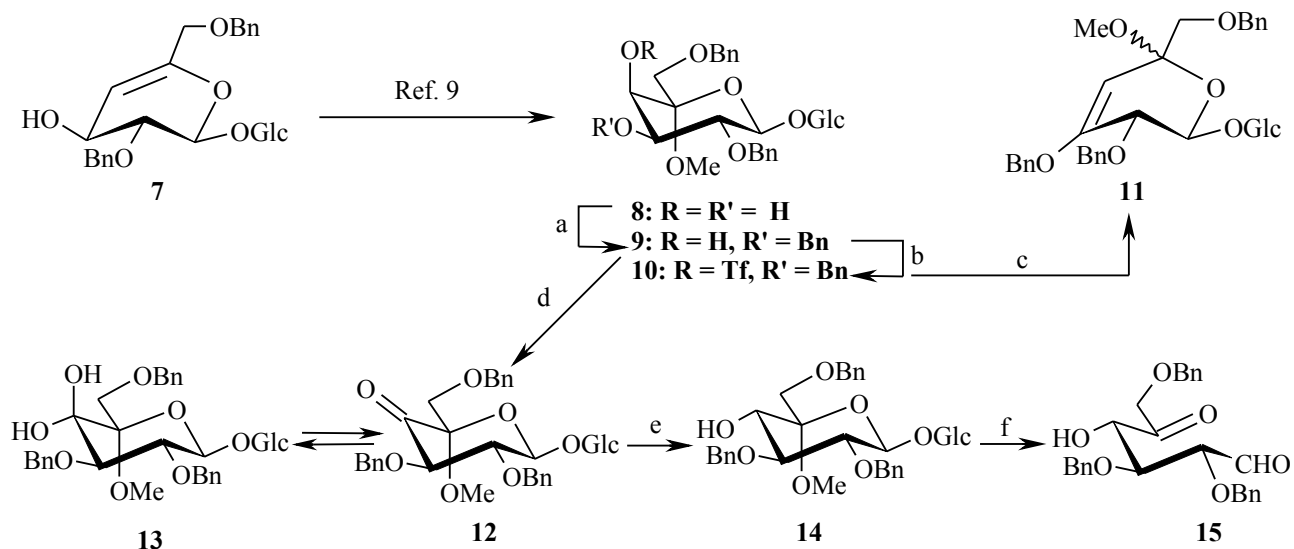


Chart 1. General approach to aldohexos-5-uloses from β -D-galactopyranosides

In this communication we present the synthesis of partially protected derivatives of D-xylo and L-lyxo-aldohexos-5-uloses achieved here from the disaccharide 1',5'-bis-glycoside **8**, analogous to **4**, in turn easily obtained from lactose,⁹ following the same general approach outlined in Chart 1. The influence of the axial C-5'-OMe group on the chemo- and the stereoselectivity of some reactions performed on the bis-glycoside unit is also observed and discussed.

2. Results and Discussion

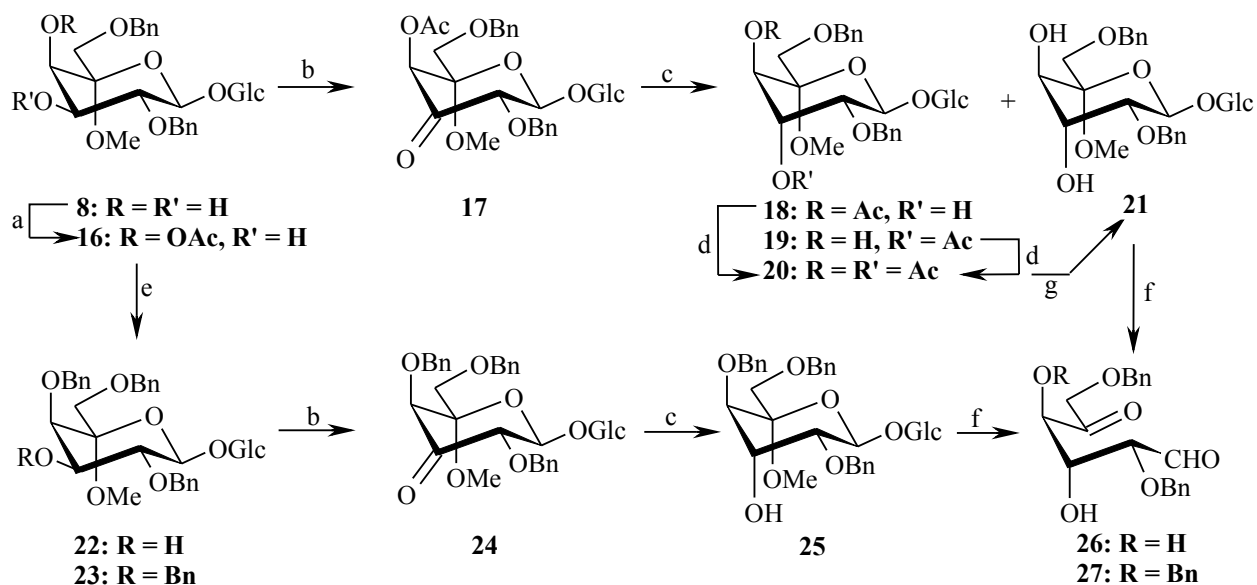
Preliminary attempts to obtain 1,5-bis-glycopyranosides of the D-xylo series through epoxidation-methanolysis of a 3'-O-protected derivative of the known disaccharide hex-4'-enopyranoside **7**,⁹ following the method previously used in monosaccharide series,^{8b} were abandoned due to the difficulties encountered in the separation of the complex crude diastereoisomeric mixture. It was considered the alternative way based on the regioselective protection of 3'-OH of the known diol **8**,⁹ followed by stereoselective epimerization at C-4' (Scheme 1). The first step was easily achieved through the stannylidene acetal-mediated alkylation method, largely used to differentiate 1,2-*cis*-diols of sugar,¹⁰ but until now, never reported on a 1,5-bis-glycopyranoside. As in the case of β -D-galactopyranosides¹⁰ the alkylation took place with complete regioselectivity on 3'-OH leading in almost quantitative yield to the alcohol **9**.



Scheme 1. Stereoselective synthesis of 2,3,6-tri-*O*-benzyl-*D*-xylo-hexos-5-ulose. Reagent and conditions: (a) Bu_2SnO , $\text{C}_6\text{H}_5\text{CH}_3$, reflux, 12 h, then BnBr , Bu_4NBr , reflux, 1.5 h (94%); (b) Tf_2O , 1:1 CH_2Cl_2 -Py, room temp, 6 h (88%); (c) Bu_4NNO_2 , $\text{C}_6\text{H}_5\text{CH}_3$, reflux, 8 h (70%); (d) TPAP, NMO, CH_2Cl_2 , 4Å, room temp, 4 h; (e) NaBH_4 , MeOH, room temp, 1.5 h, (64% from **9**); (f) 90% aq CF_3COOH , 4:1 CH_3CN - H_2O , 50 °C, 12 h (72%).

The first C-4' epimerization strategy was attempted via a $\text{S}_{\text{N}}2$ displacement on the triflate **10**, obtained from **9** in high isolated yield (88%) by treatment with Tf_2O in pyridine. Surprisingly, the treatment of **10** with Bu_4NNO_2 in toluene led to the formation of enol ether **11**, isolated in 70% yield, instead to the desired inverted bis-glycoside **14**. This result is quite unexpected in light of the high yields reported for nucleophilic substitutions of structurally related 4-*O*-triflates of galactopyranosides¹¹ and is evidently due to some interference between the axial 5'-OMe group and the nucleophile approaching the vicinal reacting centre. The complementary strategy based on an oxidation-reduction sequence of **9** was thus explored. Also in the oxidation of **9**, the presence of the axial C-5' methoxyl group sensibly influenced the reaction. Swern oxidation attempts failed completely, while the treatment with PCC showed a low conversion even after long reaction times. Better results were obtained with the TPAP-NMO system, employing however an unexpectedly high catalyst molar ratio (40%) with respect to that usually needed (5%).¹² NMR analysis of the crude oxidation product showed a mixture of the 4'-ulosyl derivative **12** (C-4' ^{13}C chemical shift: 198.3 ppm) and of its hydrate **13** (C-4' ^{13}C chemical shift: 97.1 ppm). In the crude oxidation mixture, isolated in about 89%, compounds **12** and **13** were present in about 4:1 ratio, as determined on the basis of the relative intensity of the ^{13}C 5'-OMe signals. Chromatographic purification led again to a mixture of **12** and **13** in the same 4:1 ratio, but with substantial loss of product, lowering the yield to a modest 56%. The reduction of the crude oxidation mixture with NaBH_4 in MeOH led to **14** and **9** in 64 and 20% isolated yield, respectively, indirectly confirming the structures of **12** and **13** and the extensive loss of the uloside during the chromatography on silica gel. In the case of the hydride reduction, the presence of the axial 5'-OMe group was beneficial for

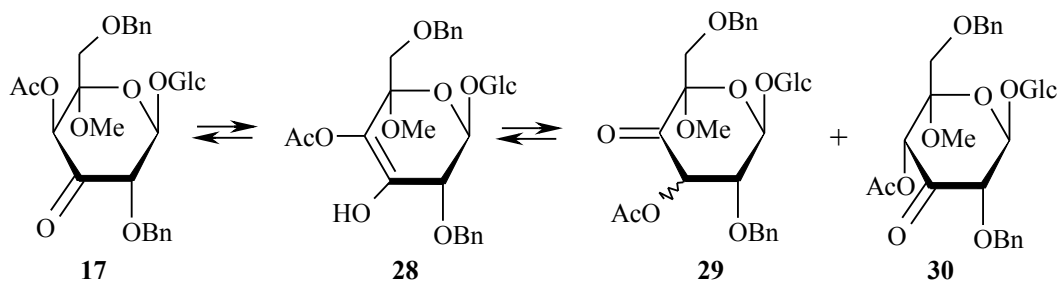
the stereoselectivity, determining the prevalence, although not complete, of attack on the β face. This result is at variance with respect to the hydride reduction of analogous 4-keto-D-*arabino*-hexopyranosides, leading mainly to D-galactopyranosides.¹³ Finally the target 2,6-di-*O*-benzyl-D-*xylo*-aldohexos-5-ulose (**15**) was obtained from **14** (72% yield) by acid hydrolysis with CF₃COOH in CH₃CN-water (50 °C, 12 h) and separation from D-glucose by extraction with EtOAc. As previously reported,^{8b} **15** was present in CD₃CN as a 55:45 α,β -1,4-furanose mixture, as confirmed by NMR analysis.



Scheme 2. Synthesis of 2,6-di-*O*-benzyl- and 2,4,6-tri-*O*-benzyl-*L*-*xylo*-aldohexos-5-ulose. Reagents and conditions: (a) CH₃C(OEt)₃, TsOH, C₆H₅CH₃, 45 °C, 25 min, then 80% aq AcOH, room temp, 15 min (quantitative); (b) TPAP, NMO, CH₂Cl₂, 4Å, room temp, 45 min (**17**: 98%, **24**: 76%); (c) NaBH₄, MeOH, room temp, 20 min (**25**: 88%); (d) 1:2 Ac₂O-Py, room temp, 20 h; (e) NaH, BnBr (1 eq), DMF, 0 °C, 25 min (**22**: 75% + **23**: 15%); (f) 90% aq CF₃COOH, 4:1 CH₃CN-H₂O, 50 °C, 4 h (**26**: 78%, **27**: 86%); (g) 0.1 M MeONa-MeOH, room temp, 2 h (quantitative).

The preparation of *L*-*xylo* derivatives was based on the same approach used in the monosaccharide series,^{8c} providing an oxidation-reduction sequence of a 3-OH free 1,5-bis-methyl *L*-*arabino*-hexopyranoside, obtained through the completely regioselective orthoester-mediated 4'-*O*-acetylation of the diol **8** (Scheme 2). As expected, the treatment of **8** with CH₃C(OEt)₃ and TsOH in toluene followed by the opening of the ortho-acetate ring with AcOH, afforded in almost quantitative yield the alcohol **16**. The next oxidation step was achieved again using the TPAP-NMO system under milder reaction conditions with respect to those used before (TPAP 5%, 2 h, 98% yield). The reduction of **17** (NaBH₄, MeOH) appeared not as simple as for the analogous 1,5-bis-methylglycopyranoside, for which only the formation of the two 5'-OMe *L*-*xylo* and *L*-*arabino* diastereoisomers in a 5:1 ratio was reported.^{8c} In fact, the reduction of the uloside **17** led to a complex product mixture, constituted (TLC, 1:1 EtOAc-hexane) of at least four components displaying two well differentiated ranges of *R*_f values on silica gel. The flash chromatography permitted only a partial separation of the two faster moving components (*R*_f 0.39 and 0.34,

respectively), surprisingly identified (NMR) as the two isomeric *L*-xylo monoacetates **18** and **19**, accounting for a combined 52% yield. The structure of the above compounds was further confirmed through conventional acetylation leading, in both cases, to the same diacetate **20**. Two other fractions containing the lower moving components (R_f 0.23 and 0.20) were collected, one constituted by the pure *L*-xylo diol **21** (13% yield) and the other by a mixture (about 1:1, combined yield 26%) of **18** and one other, yet unidentified,[§] diastereoisomeric diol. While the presence of the 4'-*O*-acetate **18** could be simply explained by a lower base-promoted *O*-deacetylation rate with respect to the monosaccharide analogue,^{8c} it is difficult to imagine the formation of the 3'-*O*-acetate **19** directly from **18** through an acetyl shift taking place, after the reduction, from the axial OH-4' group to the axial *anti* 3'-OH one. A better hypothesis as why **19** is formed would be an acyl shift on an enolic intermediate such as **28**, where the 3'-OH group and the 4'-OAc one are co-planar (Scheme 3). The enolization of **17** could explain both the migration of the acetyl group and the loss of stereochemical purity either at C-4' and C-3', giving rise to the isolation in low amount also of a unidentified diastereoisomeric diol.



Scheme 3. Base-promoted isomerization of the uloside **17**

The *O*-deacetylation of **20** (MeONa-MeOH) afforded quantitatively **21**, raising its overall yield to an acceptable 65%. Diol **21** was finally subjected to acid hydrolysis (CF₃COOH-CH₃CN-water) to give the previously reported 2,6-di-*O*-benzyl-*L*-xylo-aldohexos-5-ulose (**26**).^{8c}

The problems encountered in the reduction of **17** could be avoided simplifying the chromatographic purifications, by changing the protection of the 4'-OH group from the acetate to a base-stable ethereal one. The direct monobenylation of **8** was considered as the method of choice for the 4'-regioselective protection on the basis of the findings from Bernet and Vasella,¹⁴ which underlined, both in sugars and inositols, an enhanced acidity for axial hydroxy groups having a vicinal *anti* alkoxy group. The benzylation with BnBr (1 equiv) and NaH in DMF at 0°C pleasantly led to the 4'-*O*-monobenzylated derivative **22**, isolated in a satisfactory 75% yield, after an easy chromatographic separation from the 3,4-di-*O*-benzyl derivative **23** (Scheme 2). No traces of the product of 3'-*O*-monobenylation (**14**) were observed at any stage of the reaction. A specific role of

[§]Unexpectedly, this second diastereoisomeric diol did not correspond to the *L*-arabino derivative **8**. Owing to the difficulties to obtain a pure sample of this compound, present in the crude reaction mixture in low yield (about 13%), we abandoned any effort to further elucidate its structure, a point that was out of the scope of the present work.

the axial 5'-OMe in enhancing the acidity of 4'-OH is outlined by comparing the lower regioselectivity in the mono-benylation of methyl 2,6-di-*O*-benzyl- β -*D*-galactopyranoside where 3-OH and 4-OH alkylation products were obtained in a 1:2 ratio.¹⁵ The alcohol **22** was then submitted to an oxidation reaction with TPAP-NMO and the desired 3'-ulosyl derivative **24** was obtained in 76% yield. The reduction of **24** with NaBH₄ in MeOH afforded **25** in a completely stereoselective way, owing to the negative 1,3-*syn* di-axial interaction between the 5'-OMe group and the hydride attacking from the α face. 2,4,6-Tri-*O*-benzyl-*L*-*lyxo* aldohexos-5-ulose **27** was obtained after removal of the acetal groups by acid hydrolysis (CF₃COOH, CH₃CN-water) and its NMR data were identical to those of the sample previously reported,^{6b} pointing to a complex tautomeric mixture, whose structure was indirectly confirmed by its transformation into the expected inosose.^{6b}

In conclusion, with this work the understanding of lactose potentiality as starting renewable material for the synthesis of fine chemicals has been increased preparing aldohexos-5-ulose derivatives of the *D*-*xylo* and *L*-*lyxo* series. The crucial role of the axial anomeric 5'-OMe group of the 1',5'-bis-glycosides has also been pinpointed in both the synthetic routes. In the first one, which leads to the *D*-*xylo* derivative **15**, it decreases the reactivity in the C-4' oxidation, and moreover turns completely the reactivity of the axial 4'-*O*-trifluoromethansulfonate from the substitution to the elimination, and, for steric reasons, inverts the outcome of the 4'-uloside reduction diastereoselection from an about complete *galacto* to a prevalent *gluco*. In the second synthetic pathway, leading to the *L*-*lyxo* hexos-5-ulose **27**, the presence of the axial C-5' methoxyl group allows the regioselective alkylation of 4'-OH group and the completely stereoselective reduction of the intermediate 3'-uloside.

3. Experimental

3.1. General methods

General methods are those reported in Ref. 16. Compound **8** was prepared according to the described procedure.⁹

3.2. (5*R*)-2,3,6-Tri-*O*-benzyl-5-*C*-methoxy- α -*L*-*arabino*-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-*aldehydo*-*D*-glucose dimethyl acetal (**9**)

A soln of **8**⁹ (3.94 g, 5.80 mmol) in toluene (90 mL) was treated with Bu₂SnO (1.86 g, 7.44 mmol), heated to reflux and subjected to azeotropic removal of water with a Dean-Stark apparatus. The reaction mixture was stirred at reflux (12 h) and then treated with Bu₄NBr (944 mg,

2.90 mmol), BnBr (0.94 mL, 7.88 mmol) and further stirred until the starting material was disappeared (1.5 h, TLC, 7:3 hexane-EtOAc). The solvent was removed under diminished pressure and the residue (9.02 g) was subjected to flash chromatography (first hexane 400 mL, then 7:3 hexane-EtOAc) to give **9** (4.20 g, 94% yield) as a colourless syrup; $[\alpha]_D^{25} +4.0$ (*c* 1.2, CHCl₃); R_f 0.29 (7:3 hexane-EtOAc); ¹H NMR (600 MHz, CDCl₃): see Table 1 and δ 7.35-7.26 (m, 15H, Ar-H), 4.84, 4.68 (AB system, 2H, $J_{A,B}$ 11.1 Hz, CH₂Ph), 4.72, 4.47 (AB system, 2H, $J_{A,B}$ 12.3 Hz, CH₂Ph), 4.68 (s, 2H, CH₂Ph), 4.49 (dd, 1H, $J_{1,2}$ 6.5 Hz, $J_{2,3}$ 7.7 Hz, H-2), 4.31 (d, 1H, H-1), 4.26 (bq, 1H, H-5), 4.14 (dd, 1H, $J_{5,6b}$ 5.5 Hz, $J_{6a,6b}$ 8.9 Hz, H-6b), 4.02 (dd, 1H, $J_{3,4}$ 1.0 Hz, H-3), 3.88 (m, 1H, H-6a), 3.86 (dd, 1H, $J_{4,5}$ 5.8 Hz, H-4), 3.27, 3.26, 3.23 (3s, each 3H, 3 × OMe), 2.48 (d, 1H, $J_{4,OH}$ 1.9 Hz, OH-4'), 1.40, 1.38, 1.37, 1.36 (4s, each 3H, 2 × CMe₂); ¹³C NMR (50 MHz, CDCl₃): see Table 2 and δ 138.6, 138.0, 137.7 (3 × Ar-C), 128.4-127.4 (Ar-CH), 109.9, 108.5 (2 × CMe₂), 75.2, 73.5, 72.5 (3 × CH₂Ph), 55.7, 52.5 (2 × OMe-1), 47.9 (OMe-5'), 27.3, 26.9, 26.5, 25.3 (2 × CMe₂). Anal. Calcd for C₄₂H₅₆O₁₃: C, 65.61; H, 7.34. Found: C, 65.70; H, 7.39.

3.3. (5*R*)-2,3,6-Tri-*O*-benzyl-4-*O*-trifluoromethanesulfonyl-5-*C*-methoxy- α -*L*-arabino-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehydo-*D*-glucose dimethyl acetal (**10**)

To a soln of **9** (213 mg, 0.277 mmol) in dry 1:1 Py-CH₂Cl₂ (3 mL) cooled to -14 °C was added drop wise Tf₂O (55 μ L, 0.333 mmol) dissolved in dry CH₂Cl₂ (10 mL). The mixture was warmed to room temp and stirred until the starting material was disappeared (6 h, TLC, 6:4 hexane-EtOAc). 8 mL of satd aq NaHCO₃ were added and the mixture partitioned between water and CH₂Cl₂. The aq phase was extracted with CH₂Cl₂ (3 × 25 mL), the organic extracts collected, dried (MgSO₄) and concentrated under diminished pressure. The residue (264 mg) was subjected to flash chromatography (3:1 hexane-EtOAc) to give **10** (0.221 g, 88% yield) as a colourless syrup; $[\alpha]_D^{25} +15.7$ (*c* 1.2, CHCl₃); R_f 0.29 (7:3 hexane-EtOAc); ¹H NMR (250 MHz, CD₃CN): see Table 1 and δ 7.59-7.27 (m, 15H, Ar-H), 4.83, 4.59 (AB system, 2H, $J_{A,B}$ 11.3 Hz, CH₂Ph), 4.79, 4.67 (AB system, 2H, $J_{A,B}$ 11.4 Hz, CH₂Ph), 4.63, 4.49 (AB system, 2H, $J_{A,B}$ 11.5 Hz, CH₂Ph), 4.36 (m, 2H, H-1, H-2), 4.24 (bq, 1H, H-5), 4.08 (dd, 1H, $J_{5,6b}$ 5.7 Hz, $J_{6a,6b}$ 8.6 Hz, H-6b), 4.03 (m, 1H, $J_{3,4}$ 1.7 Hz, H-3), 3.88 (dd, 1H, $J_{4,5}$ 5.9 Hz, H-4), 3.87 (dd, 1H, $J_{5,6a}$ 5.4 Hz, H-6a), 3.32 (s, 6H, 2 × OMe-1), 3.22 (s, 3H, OMe-5'), 1.36, 1.35, 1.33, 1.26 (4s, each 3H, 2 × CMe₂); ¹³C NMR (62.9 MHz, CD₃CN): see Table 2 and δ 139.2, 138.6, 138.4 (3 × Ar-C), 129.3-128.5 (Ar-CH), 110.3, 109.2 (2 × CMe₂), 100.3 (C-1'), 75.9, 73.8, 73.7 (3 × CH₂Ph), 56.5, 53.6 (2 × OMe-1), 49.3 (OMe-5'), 27.4, 26.9, 26.7, 25.3 (2 × CMe₂). Anal. Calcd for C₄₂H₅₃F₃O₁₄S: C, 57.92; H, 6.13; F, 6.54; S, 3.68. Found: C, 57.94; H, 6.14; F, 6.56; S, 3.69.

3.4. (5*R*)-2,3,6-Tri-*O*-benzyl-4-deoxy-5-*C*-methoxy- β -*D*-glycero-hex-3-enopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehydo-*D*-glucose dimethyl acetal (**11**)

A soln of **10** (102 mg, 0.113 mmol) in dry toluene (5 mL) was treated at room temp with Bu₄NNO₂ (147 mg, 0.51 mmol) and warmed to reflux under stirring. After 8 h when the starting material was disappeared (TLC, 3:2 hexane-EtOAc) the mixture was cooled to room temp and concentrated under diminished pressure. The residue (160 mg) was subjected to flash chromatography (4:1 hexane-EtOAc), to give **11** (60 mg, 70% yield) as a colourless syrup; $[\alpha]_D$ -24.0 (*c* 1.36, CHCl₃); *R*_f 0.29 (7:3 hexane-EtOAc); ¹H NMR (250 MHz, CD₃CN): see Table 1 and δ 7.43-7.25 (m, 15H, Ar-H), 4.90, 4.84 (AB system, 2H, *J*_{A,B} 11.9 Hz, CH₂Ph), 4.74 (s, 2H, CH₂Ph), 4.55, 4.49 (AB system, 2H, *J*_{A,B} 12.0 Hz, CH₂Ph), 4.37 (d, 1H, *J*_{1,2} 5.5 Hz, H-1), 4.34 (dd, 1H, *J*_{2,3} 6.4 Hz, H-2), 4.23 (m, 1H, H-5), 4.12 (dd, 1H, *J*_{5,6b} 1.3 Hz, *J*_{6a,6b} 8.5 Hz, H-6b), 4.06 (dd, 1H, *J*_{3,4} 1.6 Hz, H-3), 4.03 (dd, 1H, *J*_{5,6a} 6.1 Hz, H-6a), 3.88 (dd, 1H, *J*_{4,5} 5.3 Hz, H-4), 3.34, 3.30, 3.25 (3s, each 3H, 2 \times OMe-1, OMe-5'), 1.33, 1.32, 1.30, 1.26 (4s, each 3H, 2 \times CMe₂); ¹³C NMR (62.9 MHz, CD₃CN): see Table 2 and δ 139.5, 139.4, 137.8 (3 \times Ar-C), 129.4-128.6 (Ar-CH), 110.5, 109.1 (2 \times CMe₂), 74.8, 74.1, 73.0 (3 \times CH₂Ph), 56.6, 53.5 (2 \times OMe-1), 50.0 (OMe-5'), 27.5, 27.1, 26.8, 25.4 (2 \times CMe₂). Anal. Calcd for C₄₁H₅₂O₁₁: C, 68.31; H, 7.27. Found: C, 68.35; H, 7.29.

3.5. (5*R*)-2,3,6-Tri-*O*-benzyl-5-*C*-methoxy- α -*L*-threo-hex-4-ulopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehydo-*D*-glucose dimethyl acetal (**12**)

A soln of **9** (493 mg, 0.641 mmol) in dry CH₂Cl₂ (12 mL) and pre-dried 4-methylmorpholine-*N*-oxide (NMO) (123 mg, 1.05 mmol) containing 4Å powdered molecular sieves (120 mg) was stirred under argon atmosphere for 30 min at room temp. Tetrapropylammonium perruthenate (TPAP) (90 mg, 40%) was added and the resulting green mixture was stirred further 4 h at room temp. The reaction mixture was filtered through alternate paths of Celite and silica gel and extensively washed with CH₂Cl₂ and then EtOAc. The combined organic phases were concentrated under diminished pressure to give a syrup (439 mg) constituted (¹³C NMR, CDCl₃) by a mixture of **12** and **13** in a ratio of about 4:1 estimated on the basis of the relative 5'-OMe signal intensities. Flash chromatographic purification of the crude residue, eluting with 6:4 hexane-EtOAc, gave a 4:1 mixture of **12** and **13** (276 mg combined yield about 56%) as a colourless syrup; $[\alpha]_D$ +9.3 (*c* 1.1, CHCl₃); *R*_f 0.92 (9:1 CH₂Cl₂-Me₂CO); selected ¹³C NMR (50 MHz, CD₃CN) signals: *major component 12*: δ 198.3 (C-4'), 139.7, 137.4, 137.3 (3 \times Ar-C), 110.4, 109.2 (2 \times CMe₂), 106.6 (C-1), 99.4 (C-5'), 99.1 (C-1'), 84.7, 82.3 (C-2', C-3'), 78.4, 77.9 (C-3, C-5), 76.7, 75.9 (C-4, C-2),

75.7, 75.7, 74.1 ($3 \times \text{CH}_2\text{Ph}$), 67.2, 66.0 (C-6, C-6'), 56.6, 53.9 ($2 \times \text{OMe-1}$), 50.2 (OMe-5'); *minor component 13*: δ 139.8, 138.9, 138.2 ($3 \times \text{Ar-C}$), 110.3, 109.2 ($2 \times \text{CMe}_2$), 106.4 (C-1), 99.4 (C-5'), 99.4 (C-1'), 97.1 (C-4'), 82.5, 81.5 (C-2', C-3'), 81.4, 78.4 (C-3, C-5), 77.6, 77.5 (C-4, C-2), 76.4, 75.7, 74.1 ($3 \times \text{CH}_2\text{Ph}$), 70.3, 66.0 (C-6, C-6'), 56.4, 53.6 ($2 \times \text{OMe-1}$), 48.8 (OMe-5'). Cluster of signals for both components: δ 129.4-127.5 (Ar-CH), 27.8-25.4 (CMe_2).

3.6. (5*R*)-2,3,6-Tri-*O*-benzyl-5-*C*-methoxy- β -*D*-xylo-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehydo-*D*-glucose dimethyl acetal (**14**)

A soln of the crude **12** and **13** mixture (912 mg) in dry MeOH (50 mL) was cooled at 0 °C and treated with NaBH₄ (180 mg, 4.77 mmol). The reaction mixture was gently warmed to room temp and left under stirring until the TLC analysis (7:3 hexane-EtOAc) showed the complete disappearance of the starting material (1.5 h). Water (15 mL) was added, the soln stirred for 30 min, concentrated under diminished pressure and the residue partitioned between water (40 mL) and CH₂Cl₂ (80 mL). The aq phase was extracted with CH₂Cl₂ (3×80 mL), the organic extracts were collected, dried (MgSO₄), concentrated under diminished pressure and the residue (869 mg) subjected to flash chromatography over silica gel (7:3 hexane-EtOAc) to give **14** (583 mg, 64% yield) and **9** (187 mg, 20% yield).

Compound **14**: colourless syrup; $[\alpha]_D$ -15.9 (c 1.0, CHCl₃); R_f 0.12 (7:3 hexane-EtOAc); ¹H NMR (200 MHz, CD₃CN): see Table 1 and δ 7.39-7.26 (m, 15H, Ar-H), 4.84, 4.66 (AB system, 2H, $J_{A,B}$ 11.4 Hz, CH₂Ph), 4.81, 4.75 (AB system, 2H, $J_{A,B}$ 11.3 Hz, CH₂Ph), 4.62, 4.48 (AB system, 2H, $J_{A,B}$ 11.8 Hz, CH₂Ph), 4.48 (dd, 1H, $J_{1,2}$ 6.3 Hz, $J_{2,3}$ 7.4 Hz, H-2), 4.33 (d, 1H, H-1), 4.21 (m, 1H, H-5), 4.09 (dd, 1H, $J_{5,6b}$ 5.7 Hz, $J_{6a,6b}$ 8.5 Hz, H-6b), 4.08 (dd, 1H, $J_{3,4}$ 1.6 Hz, H-3), 3.87 (dd, 1H, $J_{5,6a}$ 6.2 Hz, H-6a), 3.85 (bd, 1H, H-4), 3.36, 3.32, 3.30 (3s, each 3H, $2 \times \text{OMe-1}$, OMe-5'), 1.36, 1.35, 1.34, 1.26 (4s, each 3H, $2 \times \text{CMe}_2$); ¹³C NMR (50 MHz, CD₃CN): see Table 2 and δ 140.2, 139.7, 139.2 ($3 \times \text{Ar-C}$), 129.3-128.2 (Ar-CH), 110.4, 109.2 ($2 \times \text{CMe}_2$), 75.7, 75.5, 74.3 ($3 \times \text{CH}_2\text{Ph}$), 56.6, 54.3 ($2 \times \text{OMe-1}$), 49.0 (OMe-5'), 27.5, 27.1, 27.0, 25.5 ($2 \times \text{CMe}_2$). Anal. Calcd for C₄₂H₅₆O₁₃: C, 65.61; H, 7.34. Found: C, 65.73; H, 7.38.

Compound **9**: colourless syrup; NMR parameters were identical to those of the sample prepared above.

3.7. 2,3,6-Tri-*O*-benzyl-*D*-xylo-hexos-5-ulose (**15**)

A soln of **14** (276 mg, 0.359 mmol) in 4:1 (v/v) CH₃CN-water (7 mL) was treated with 90% aq CF₃COOH (1.4 mL) warmed to 50 °C and stirred until the TLC analysis (EtOAc) showed the

complete disappearance of the starting material (12 h). The mixture was concentrated under diminished pressure and repeatedly co-evaporated with toluene (4×20 mL) under diminished pressure. The crude residue was partitioned between brine (20 mL) and EtOAc (30 mL) and the aq phase extracted with EtOAc (3×30 mL). The organic phases were collected, dried (MgSO_4), concentrated under diminished pressure to give a residue (125 mg), that was directly filtered on silica gel eluting with 3:7 hexane-EtOAc, to give pure **15** (122 mg, 72% yield) as colourless syrup. NMR data were in full agreement with the reported ones.^{8b}

3.8. (5*R*)-4-*O*-Acetyl-2,6-di-*O*-benzyl-5-*C*-methoxy- α -L-arabino-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehydo-D-glucose dimethyl acetal (**16**)

To a soln of **8**⁹ (341 mg, 0.502 mmol) in dry toluene (10 mL) warmed to 45 °C, $\text{CH}_3\text{C}(\text{OEt})_3$ (1.1 mL, 6.02 mmol) and TsOH (9.5 mg, 0.05 mmol) were added. The soln was stirred for 2 h at 45 °C until the TLC analysis (1:1 hexane-EtOAc) showed the complete disappearance of the starting material (R_f 0.38) and the formation a faster moving product (R_f 0.60). The mixture was allowed to obtain room temp, treated with Et_3N (0.1 mL) and further stirred for 10 min. The soln was concentrated at diminished pressure and the residue (228 mg) was treated with 80% aq AcOH (3.0 mL) and stirred at room temp until the product at R_f 0.60 was completely reacted (TLC, 1:1 hexane-EtOAc, 15 min). The mixture was diluted with CH_2Cl_2 (20 mL) and carefully neutralized with 40% aq NaOH (4 mL). The reaction mixture was diluted with water (7 mL) and the aq phase was extracted with CH_2Cl_2 (3×20 mL). The combined organic layers were dried (MgSO_4), filtered, and concentrated under diminished pressure. The residue (356 mg) was constituted exclusively (NMR) of **16** (quantitative yield). An analytical sample of **16** was obtained through flash chromatography eluting with 3:2 hexane-EtOAc. Pure **16** (308 mg, 85% yield) was a white foam; $[\alpha]_D -24.2$ (c 0.95, CHCl_3); R_f 0.51 (1:1 hexane-EtOAc); ^1H NMR (200 MHz, CDCl_3): see Table 1 and δ 7.35-7.26 (m, 10H, Ar-H), 4.92, 4.59 (AB system, 2H, $J_{A,B}$ 11.3 Hz, CH_2Ph), 4.49 (dd, 1H, $J_{1,2}$ 6.7 Hz, $J_{2,3}$ 7.0 Hz, H-2), 4.46 (s, 2H, CH_2Ph), 4.32 (d, 1H, H-1), 4.30-3.95 (m, 5H, H-3, H-4, H-5, H-6a, H-6b), 3.34, 3.33 (2s, each 3H, $2 \times \text{OMe-1}$), 3.27 (s, 3H, OMe-5'), 2.22 (bs, 1H, OH-3'), 1.96 (s, 3H, MeCO), 1.45, 1.32 (2s, each 3H, CMe_2), 1.41 (s, 6H, CMe_2); ^{13}C NMR (50 MHz, CDCl_3): δ see Table 2 and 169.5 (MeCO), 137.8, 136.9 ($2 \times \text{Ar-C}$), 129.4-127.4 (Ar-CH), 109.6, 108.1 ($2 \times \text{CMe}_2$), 74.3, 73.1 ($2 \times \text{CH}_2\text{Ph}$), 55.6, 52.9 ($2 \times \text{OMe-1}$), 47.8 (OMe-5'), 26.8, 26.1, 26.0, 24.7 ($2 \times \text{CMe}_2$), 20.4 (MeCO). Anal. Calcd for $\text{C}_{37}\text{H}_{52}\text{O}_{14}$: C, 61.65; H, 7.27. Found: C, 61.59; H, 7.23.

3.9. Oxidation-reduction of (5*R*)-4-*O*-Acetyl-2,6-di-*O*-benzyl-5-*C*-methoxy- α -L-arabino-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehydo-D-glucose dimethyl acetal (**16**)

3.9.1. Oxidation of 16. - A suspension of **16** (356 mg, 0.494 mmol) in dry CH₂Cl₂ (6 mL) and pre-dried 4-methylmorpholine-*N*-oxide (NMO) (98 mg, 0.841 mmol) containing 4Å powdered molecular sieves (250 mg) was stirred under argon atmosphere for 30 min at room temp. Tetrapropylammonium perruthenate (TPAP) (35 mg, 20%) was added and the resulting green mixture was stirred until the TLC analysis (13:7 hexane-EtOAc) revealed the disappearance of the starting material (45 min, *R_f* 0.25). The reaction mixture was filtered through alternate paths of Celite and silica gel and extensively washed first with CH₂Cl₂ and then with EtOAc. The combined organic phases were concentrated under diminished pressure to give a syrup (352 mg) constituted exclusively (NMR) by the uloside **17**; *R_f* 0.17 (13:7 hexane-EtOAc); ¹H NMR (200 MHz, CDCl₃): see Table 1 and δ 7.30-7.22 (m, 10H, Ar-H), 4.80, 4.65 (AB system, 2H, *J_{A,B}* 11.9 Hz, CH₂Ph), 4.52, 4.43 (AB system, 2H, *J_{A,B}* 12.4 Hz, CH₂Ph), 4.46 (dd, 1H, *J_{1,2}* 6.6 Hz, *J_{2,3}* 7.1 Hz, H-2), 4.32 (d, 1H, H-1), 4.25 (m, 1H, H-5), 4.17-3.85 (m, 4H, H-3, H-4, H-6a, H-6b), 3.36 (s, 6H, 2 × OMe-1), 3.29 (s, 3H, OMe-5'), 1.86 (s, 3H, MeCO), 1.41, 1.38, 1.35, 1.31 (4s, each 3H, 2 × CMe₂); ¹³C NMR (50 MHz, CDCl₃): see Table 2 and δ 168.5 (MeCO), 137.1, 136.8 (2 × Ar-C), 129.7-127.8 (Ar-CH), 109.9, 108.6 (2 × CMe₂), 73.2, 72.9 (2 × CH₂Ph), 56.2, 53.6 (2 × OMe-1), 48.7 (OMe-5'), 27.2, 26.5, 26.3, 25.3 (2 × CMe₂), 20.3 (MeCO).

3.9.2. Reduction of 17. - A soln of crude **17** (352 mg, 0.489 mmol) in dry MeOH (10 mL) was cooled to 0 °C and treated, under argon atmosphere, with NaBH₄ (138 mg, 3.65 mmol). The reaction mixture was gently warmed to room temp and left under stirring until the TLC analysis (1:1 hexane-EtOAc) showed the complete disappearance of the starting material (30 min). Water (15 mL) was added, the soln was stirred for 4 h, concentrated under diminished pressure and the residue partitioned between water (20 mL) and CH₂Cl₂ (40 mL). The aq phase was extracted with CH₂Cl₂ (5 × 25 mL) and the organic extracts were collected, dried (MgSO₄) and concentrated under diminished pressure. The residue (307 mg) was subjected to flash chromatography (first 11:9 hexane-EtOAc, then 2:3 hexane-EtOAc) collecting four main fractions. The first two fractions were constituted by the monoacetates **18** (65 mg) and **19** (120 mg), each in mixture with about 10% of the other, accounting for an overall 52% yield, the third fraction contained pure diol **21** (43 mg, 13% yield), and the fourth one an about 1:1 mixture of **21** and another yet unidentified diastereoisomeric diol (86 mg, each about 13% yield).

(5R)-4-O-Acetyl-2,6-di-O-benzyl-5-C-methoxy-α-L-xylo-hexopyranosyl-(1 → 4)-2,3:5,6-di-O-isopropylidene-aldehydo-D-glucose dimethyl acetal (18): *R_f* 0.39 (1:1 hexane-EtOAc); ¹H NMR (200 MHz, CDCl₃): see Table 1 and δ 7.34-7.18 (m, 10H, Ar-H), 4.71, 4.59 (AB system, 2H, *J_{A,B}* 12.0 Hz, CH₂Ph), 4.56, 4.49 (AB system, 2H, *J_{A,B}* 12.0 Hz, CH₂Ph), 4.42 (dd, 1H, *J_{1,2}* 6.4 Hz, *J_{2,3}* 7.0 Hz, H-2), 4.32 (d, 1H, H-1), 4.25 (m, 2H, H-5, H-6a), 4.05-3.90 (m, 3H, H-3, H-4, H-6b), 3.32,

3.30, 3.29 (3s, each 3H, 2 × OMe-1, OMe-5'), 2.69 (d, 1H, $J_{4',OH}$ 5.3 Hz, OH-4'), 2.02 (s, 3H, MeCO), 1.41, 1.32 (2s, each 3H, CMe₂); 1.40 (s, 6H, CMe₂); ¹³C NMR (50 MHz, CDCl₃): see Table 2 and δ 170.5 (MeCO), 138.2, 137.4 (2 × Ar-C), 128.5-126.9 (Ar-CH), 110.0, 108.4 (2 × CMe₂), 73.4, 72.6 (2 × CH₂Ph), 55.8, 52.8 (2 × OMe-1), 48.1 (OMe-5'), 27.3, 26.6, 26.3, 25.4 (2 × CMe₂), 20.9 (MeCO). Anal. Calcd for C₃₇H₅₂O₁₄: C, 61.65; H, 7.27. Found: C, 61.58; H, 7.25.

(5*R*)-3-*O*-Acetyl-2,6-di-*O*-benzyl-5-*C*-methoxy- α -*L*-xylo-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehyde-*D*-glucose dimethyl acetal (**19**): R_f 0.34 (1:1 hexane-EtOAc); ¹H NMR (200 MHz, CDCl₃): see Table 1 and δ 7.33-7.26 (m, 10H, Ar-H), 4.81, 4.63 (AB system, 2H, $J_{A,B}$ 12.1 Hz, CH₂Ph), 4.47, 4.40 (AB system, 2H, $J_{A,B}$ 11.7 Hz, CH₂Ph), 4.47 (dd, 1H, $J_{1,2}$ 6.4 Hz, $J_{2,3}$ 7.6 Hz, H-2), 4.28-3.94 (m, 3H, H-5, H-6a, H-6b), 4.32 (d, 1H, H-1), 4.06 (dd, 1H, $J_{3,4}$ 1.2 Hz, H-3), 3.92 (dd, 1H, $J_{4,5}$ 5.0 Hz, H-4), 3.37, 3.36, 3.35 (3s, each 3H, 2 × OMe-1, OMe-5'), 3.14 (d, 1H, $J_{3',OH}$ 8.0 Hz, OH-3'), 1.78 (s, 3H, MeCO), 1.43 (s, 6H, CMe₂); 1.42, 1.32 (2s, each 3H, CMe₂); ¹³C NMR (50 MHz, CDCl₃): see Table 2 and δ 168.5 (MeCO), 137.9, 137.3 (2 × Ar-C), 128.3-127.6 (Ar-CH), 109.9, 108.5 (2 × CMe₂), 73.3, 72.5 (2 × CH₂Ph), 56.1, 53.7 (2 × OMe-1), 48.4 (OMe-5'), 27.2, 26.5, 26.4, 25.3 (2 × CMe₂), 20.5 (MeCO). Anal. Calcd for C₃₇H₅₂O₁₄: C, 61.65; H, 7.27. Found: C, 61.60; H, 7.22.

(5*R*)-2,6-di-*O*-benzyl-5-*C*-methoxy- α -*L*-xylo-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehyde-*D*-glucose dimethyl acetal (**21**): white foam; $[\alpha]_D$ -27.2 (c 0.99, CHCl₃); R_f 0.32 (2:3 hexane-EtOAc); ¹H NMR (250 MHz, CDCl₃): δ 7.31-7.22 (m, 10H, Ar-H), 5.18 (d, 1H, $J_{1',2'}$ 7.8 Hz, H-1'), 4.81, 4.65 (AB system, 2H, $J_{A,B}$ 12.2 Hz, CH₂Ph), 4.57, 4.50 (AB system, 2H, $J_{A,B}$ 12.3 Hz, CH₂Ph), 4.47 (dd, 1H, $J_{1,2}$ 6.7 Hz, $J_{2,3}$ 7.1 Hz, H-2), 4.31 (d, 1H, H-1), 4.30-4.17 (m, 2H, H-5, H-6b), 4.05-3.98 (m, 5H, H-3', H-4', H-3, H-4, H-6a), 3.60-3.50 (m, 3H, H-2', H-6'a, H-6'b), 3.31 (s, 9H, 2 × OMe-1, OMe-5'), 3.13 (d, 1H, $J_{3',OH}$ 7.5 Hz, OH-3'), 2.60 (d, 1H, $J_{4',OH}$ 5.2 Hz, OH-4'), 1.41 (s, 9H, CMe₂); 1.32 (s, 3H, CMe₂); ¹³C NMR (62.9 MHz, CDCl₃): see Table 2 and δ 138.1, 137.2 (2 × Ar-C), 128.5-127.5 (Ar-CH), 109.9, 108.5 (2 × CMe₂), 73.3, 72.6 (2 × CH₂Ph), 56.0, 53.1 (2 × OMe-1), 48.4 (OMe-5'), 27.2, 26.6, 26.5, 25.3 (2 × CMe₂). Anal. Calcd for C₃₅H₅₀O₁₃: C, 61.93; H, 7.42. Found: C, 61.90; H, 7.39.

Selected ¹³C NMR (50 MHz, CDCl₃) data for the unidentified diastereoisomeric diol isolated in mixture with **21**: δ 106.0 (C-1), 98.6 (C-5'), 98.2 (C-1'), 81.2 (C-2'), 77.8, 77.6, 77.3 (C-3, C-4, C-5), 75.5, 74.8, 71.6 (C-2, C-3', C-4'), 73.7, 73.74 (2 × CH₂Ph), 69.9 (C-6'), 65.2 (C-6), 56.0, 53.6 (2 × OMe-1), 48.5 (OMe-5').

3.10. (5*R*)-3,4-di-*O*-Acetyl-2,6-di-*O*-benzyl-5-*C*-methoxy- α -*L*-lyxo-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehyde-*D*-glucose dimethyl acetal (**20**)

Compound **21** (40 mg, 0.059 mmol) was acetylated with a 1:2 mixture of Ac₂O and pyridine (3 mL) and stirred at room temp. After 20 h the reaction mixture was repeatedly co-evaporated with toluene (4 x 10 mL) at diminished pressure. Flash chromatographic purification (13:7 hexane-EtOAc) of the residue (52 mg) gave **20** (44 mg, 97% yield) as a syrup; $[\alpha]_D -46.9$ (*c* 0.9, CHCl₃); *R_f* 0.38 (3:2 hexane-EtOAc); ¹H NMR (200 MHz, CDCl₃): see Table 1 and δ 7.35-7.23 (m, 10H, Ar-H), 4.68, 4.58 (AB system, 2H, *J_{A,B}* 11.9 Hz, CH₂Ph), 4.43, 4.40 (s, 2H, CH₂Ph), 4.46 (dd, 1H, *J_{1,2}* 6.5 Hz, *J_{2,3}* 7.3 Hz, H-2), 4.33 (d, 1H, H-1), 4.26 (m, 1H, H-5), 4.03 (dd, 1H, *J_{3,4}* 1.3 Hz, H-3), 4.16 (dd, 1H, *J_{5,6b}* 6.2 Hz, *J_{6a,6b}* 8.5 Hz, H-6b), 3.98 (m, 2H, H-4, H-6a), 3.34, 3.33, 3.32 (3s, each 3H, 2 x OMe-1, OMe-5'), 2.00, 1.85 (2s, each 3H, 2 x MeCO), 1.41 (s, 6H, CMe₂); 1.40, 1.32 (2s, each 3H, CMe₂); ¹³C NMR (50 MHz, CDCl₃): see Table 2 and δ 169.5, 168.1 (2 x MeCO), 137.8, 137.3 (2 x Ar-C), 128.4-127.5 (Ar-CH), 110.1, 108.4 (2 x CMe₂), 73.4, 72.6 (2 x CH₂Ph), 55.8, 53.4 (2 x OMe-1), 48.1 (OMe-5'), 27.2, 26.6, 26.2, 25.4 (2 x CMe₂), 20.8, 20.6 (2 x MeCO). Anal. Calcd for C₃₉H₅₄O₁₅: C, 61.41; H, 7.14. Found: C, 61.38; H, 7.12.

The acetylation of either the fractions containing **19** (115 mg, 0.160 mmol) or **18** (65 mg, 0.090 mmol) as reported above gave, after chromatographic purification (13:7 hexane-EtOAc), **21** (183 mg, 96% yield) having NMR parameters identical to those of the sample prepared above.

3.11. (5*R*)-2,6-di-*O*-benzyl-5-*C*-methoxy- α -*L*-lyxo-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehydo-*D*-glucose dimethyl acetal (**21**)

To a soln of **20** (183 mg, 0.240 mmol) in dry MeOH (5 mL) was added at room temp a 0.1 M methanolic soln of MeONa (0.2 mL). The reaction mixture was stirred until TLC analysis (3:2 hexane-EtOAc) showed the complete disappearance of the starting material (2 h) and the formation of a lower moving product. The solution was neutralized with resin acid (Amberlyst 15) and the suspension was filtered and concentrated to give a foam residue constituted by pure (NMR) **21** (162.5 mg, quantitative yield), identical to the sample obtained above. The combined overall yield of **21**, obtained by adding the sample directly isolated in the oxidation-reduction of **16**, was 65%.

3.12. 2,6-Di-*O*-benzyl-*L*-lyxo-hexos-5-ulose (**26**)

A soln of **21** (185 mg, 0.272 mmol) in 4:1 (v/v) CH₃CN-water (6 mL) was treated with 90% aq CF₃COOH (1.2 mL) warmed to 50 °C and stirred until TLC analysis (EtOAc) showed the complete disappearance of the starting material (5 h). The mixture was concentrated under diminished pressure and repeatedly co-evaporated with toluene (5 x 20 mL). The residue was partitioned

between brine (20 mL) and EtOAc (40 mL) and the aq phase extracted with EtOAc (3 × 40 mL). The organic phases were collected, dried (MgSO₄), concentrated under diminished pressure to give a residue (96 mg), that was directly subjected to a flash chromatographic purification, eluting with 1:3 hexane-EtOAc, to give pure **26** (75 mg, 78% yield) as colourless syrup. NMR data were in agreement with the reported ones.^{8b}

3.13. (5R)-2,4,6-Tri-O-benzyl-5-C-methoxy- α -L-arabino-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-O-isopropylidene-aldehydo-D-glucose dimethyl acetal (22**) and (5R)-2,3,4,6-tetra-O-benzyl-5-C-methoxy- α -L-arabino-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-O-isopropylidene-aldehydo-D-glucose dimethyl acetal (**23**)**

A suspension of pre-washed (hexane) 60% NaH in mineral oil (1.29 g, 53.7 mmol) in dry DMF (25.0 mL) was cooled to 0 °C and treated, under argon atmosphere, with a soln of **8** (3.62 g, 5.34 mmol) in dry DMF (100 mL). The mixture was warmed to room temp and stirred for 30 min, cooled again to 0 °C and treated with BnBr (0.63 mL, 5.34 mmol) and further stirred until the starting material was consumed (25 min, TLC, 7:3 hexane-EtOAc). MeOH (12 mL) and water (100 mL) were slowly added and the reaction mixture was extracted with Et₂O (4 × 50 mL). The combined extracts were collected, dried (MgSO₄), concentrated under diminished pressure and the residue (4.68 g) subjected to flash chromatography (first hexane 600 mL, then 4:1 hexane-EtOAc) to give **22** (3.09 g, 75% yield) and **23** (684 mg, 15% yield).

Compound **22** was a colourless syrup; $[\alpha]_D^{25} +16.8$ (*c* 1.07, CHCl₃); *R_f* 0.24 (7:3 hexane-EtOAc); ¹H NMR (600 MHz, CDCl₃): see Table 1 and δ 7.36-7.22 (m, 15H, Ar-H), 4.76, 4.60 (AB system, 2H, *J_{A,B}* 11.6 Hz, CH₂Ph), 4.62, 4.58 (AB system, 2H, *J_{A,B}* 11.3 Hz, CH₂Ph), 4.58, 4.34 (AB system, 2H, *J_{A,B}* 12.1 Hz, CH₂Ph), 4.49 (dd, 1H, *J_{1,2}* 6.5 Hz, *J_{2,3}* 7.6 Hz, H-2), 4.31 (d, 1H, H-1), 4.27 (bq, 1H, H-5), 4.16 (dd, 1H, *J_{5,6b}* 5.6 Hz, *J_{6a,6b}* 8.7 Hz, H-6b), 4.00 (m, 1H, H-4), 3.95 (dd, 1H, *J_{5,6a}* 6.2 Hz, H-6a), 3.92 (dd, 1H, *J_{3,4}* 0.8 Hz, H-3), 3.27, 3.25, 3.22 (3s, each 3H, 2 × OMe-1, OMe-5'), 1.43, 1.41, 1.38, 1.31 (4s, each 3H, 2 × CMe₂); ¹³C NMR (50 MHz, CDCl₃): see Table 2 and δ 138.6, 138.3, 137.2 (3 × Ar-C), 128.3-127.2 (Ar-CH), 109.8, 108.4 (2 × CMe₂), 74.9, 74.5, 73.2 (3 × CH₂Ph), 55.6, 52.3 (2 × OMe-1), 47.9 (OMe-5'), 27.2, 26.5, 26.4, 25.0 (2 × CMe₂). Anal. Calcd for C₄₂H₅₆O₁₃: C, 65.61; H, 7.34. Found: C, 65.73; H, 7.44.

Compound **23** was a colourless syrup; $[\alpha]_D^{25} +17.6$ (*c* 1.2, CHCl₃); *R_f* 0.38 (7:3 hexane-EtOAc); ¹H NMR (200 MHz, CDCl₃): see Table 1 and δ 7.33-7.23 (m, 20H, Ar-H), 4.95, 4.57 (AB system, 2H, *J_{A,B}* 11.5 Hz, CH₂Ph), 4.83, 4.73 (AB system, 2H, *J_{A,B}* 10.7 Hz, CH₂Ph), 4.70 (s, 2H, CH₂Ph), 4.56, 4.28 (AB system, 2H, *J_{A,B}* 12.1 Hz, CH₂Ph), 4.54 (dd, 1H, *J_{1,2}* 6.7 Hz, *J_{2,3}* 9.7 Hz, H-2), 4.31 (d, 1H, H-1), 4.18 (m, 1H, H-5), 4.05 (m, 2H, H-4, H-6b), 3.98 (dd, 1H, *J_{3,4}* 2.9 Hz, H-3), 3.87 (dd,

1H, $J_{5,6a}$ 5.8 Hz, $J_{6a,6b}$ 8.9 Hz, H-6a), 3.25, 3.22, 3.21 (3s, each 3H, 2 × OMe-1, OMe-5'), 1.42, 1.41, 1.37, 1.30 (4s, each 3H, 2 × CMe₂); ¹³C NMR (50 MHz, CDCl₃): see Table 2 and δ 138.7, 138.5, 138.4, 137.2 (4 × Ar-C), 128.1-127.1 (Ar-CH), 109.6, 108.3 (2 × CMe₂), 74.8, 74.5, 73.1, 72.6 (4 × CH₂Ph), 55.4, 52.0 (2 × OMe-1), 47.8 (OMe-5'), 27.2, 26.6, 26.1, 25.1 (2 × CMe₂). Anal. Calcd for C₄₉H₆₂O₁₃: C, 68.51; H, 7.27. Found: C, 68.63; H, 7.34.

3.14. (5*R*)-2,4,6-Tri-*O*-benzyl-5-*C*-methoxy- α -*L*-threo-hex-3-ulopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehydo-*D*-glucose dimethyl acetal (**24**)

A suspension of **22** (1.03 g, 1.34 mmol) in dry CH₂Cl₂ (27 mL), pre-dried 4-methylmorpholine-*N*-oxide (NMO) (275 mg, 2.34 mmol) and 4 Å powdered molecular sieves (400 mg) was stirred under argon atmosphere for 30 min at room temp. Tetrapropylammonium perruthenate (TPAP) (47 mg, 10%) was added and the resulting green mixture was stirred for 4 h at room temp until the TLC (9:1 CH₂Cl₂-Me₂CO) showed the complete disappearance of the starting material. The reaction mixture was filtered through alternate paths of Celite and silica gel and extensively washed with CH₂Cl₂ and EtOAc. The soln and washings were combined and concentrated under diminished pressure to give almost pure **24** (NMR). A sample of the residue was subjected to flash chromatography (7:3 hexane-EtOAc) to give **24** (76% yield) as a colourless syrup; $[\alpha]_D$ -37.4 (*c* 1.16, CHCl₃); *R*_f 0.73 (9:1 CH₂Cl₂-Me₂CO); ¹H NMR (200 MHz, CD₃CN): see Table 1 and δ 7.38-7.13 (m, 15H, Ar-H), 4.70, 4.64 (AB system, 2H, $J_{A,B}$ 12.5 Hz, CH₂Ph), 4.53, 4.47 (AB system, 2H, $J_{A,B}$ 12.4 Hz, CH₂Ph), 4.40-4.32 (m, 4H, H-1, H-2, CH₂Ph), 4.21 (m, 1H, H-5), 4.07 (dd, 1H, $J_{5,6b}$ 6.0 Hz, $J_{6a,6b}$ 8.5 Hz, H-6b), 4.03 (dd, 1H, $J_{2,3}$ 6.9 Hz, $J_{3,4}$ 1.1 Hz, H-3), 3.93 (dd, 1H, $J_{5,6a}$ 6.3 Hz, H-6a), 3.87 (dd, 1H, $J_{4,5}$ 5.5 Hz, H-4), 3.31, 3.25, 3.23 (3s, each 3H, 2 × OMe-1, OMe-5'), 1.34, 1.33, 1.32, 1.27 (4s, each 3H, 2 × CMe₂); ¹³C NMR (50 MHz, CD₃CN): see Table 2 and δ 138.8, 138.6, 137.8 (3 × Ar-C), 129.3-128.8 (Ar-CH), 110.4, 109.2 (2 × CMe₂), 74.0, 73.9, 73.0 (3 × CH₂Ph), 56.5, 53.8 (2 × OMe-1), 49.1 (OMe-5'), 27.4, 26.9, 26.8, 25.5 (2 × CMe₂). Anal. Calcd for C₄₂H₅₄O₁₃: C, 65.78; H, 7.10. Found: C, 65.88; H, 7.38.

3.15. (5*R*)-2,4,6-Tri-*O*-benzyl-5-*C*-methoxy- α -*L*-lyxo-hexopyranosyl-(1 \rightarrow 4)-2,3:5,6-di-*O*-isopropylidene-aldehydo-*D*-glucose dimethyl acetal (**25**)

A soln of **24** (760 mg, 0.99 mmol) in dry MeOH (22 mL) was cooled to 0 °C and treated, under argon atmosphere, with NaBH₄ (113 mg, 2.98 mmol). After 5 min the reaction mixture was gently warmed to room temp and left under stirring until **24** was consumed (1.5 h, TLC, 9:1 CH₂Cl₂-Me₂CO). Water (15 mL) was added, the soln stirred for 30 min, concentrated under diminished

pressure and the residue partitioned between water (60 mL) and CH₂Cl₂ (150 mL). The aq phase was extracted with CH₂Cl₂ (2 × 150 mL), the organic extracts collected, dried (MgSO₄) and concentrated under diminished pressure. The residue (784 mg) was subjected to flash chromatography (6:4 hexane-EtOAc) to give **25** as a colourless syrup (668 mg, 88% yield); [α]_D +2.20 (*c* 0.7, CHCl₃); *R*_f 0.38 (9:1 CH₂Cl₂-Me₂CO); ¹H NMR (200 MHz, CDCl₃): see Table 1 and δ 7.38-7.05 (m, 15H, Ar-H), 4.81, 4.63 (AB system, 2H, *J*_{A,B} 12.2 Hz, CH₂Ph), 4.58, 4.36 (AB system, 2 H, *J*_{A,B} 12.1 Hz, CH₂Ph), 4.48 (dd, 1 H, *J*_{1,2} 6.7 Hz, *J*_{2,3} 7.6 Hz, H-2), 4.47, 4.39 (AB system, 2H, *J*_{A,B} 11.4 Hz, CH₂Ph), 4.31 (d, 1H, H-1), 4.24 (m, 1H, H-5), 4.03 (dd, 1H, *J*_{3,4} 1.0 Hz, H-3), 4.00 (m, 2H, H-6a, H-6b), 3.88 (dd, 1H, *J*_{4,5} 5.2 Hz, H-4), 3.28, 3.25, 3.15 (3s, each 3H, 2 × OMe-1, OMe-5'), 1.44, 1.43, 1.42, 1.33 (4s, each 3H, 2 × CMe₂); ¹³C NMR (50 MHz, CDCl₃): see Table 2 and δ 138.9, 138.3, 138.1 (3 × Ar-C), 128.9-128.3 (Ar-CH), 110.5, 109.2 (2 × CMe₂), 73.9, 73.7, 73.2 (3 × CH₂Ph), 56.5, 53.4 (2 × OMe-1), 48.9 (OMe-5'), 27.9, 27.2, 27.1, 26.0 (2 × CMe₂). Anal. Calcd for C₄₂H₅₆O₁₃: C, 65.61; H, 7.34. Found: C, 65.68; H, 7.41.

3.16. 2,4,6-Tri-*O*-benzyl-*L*-lyxo-hexos-5-ulose (**27**)

A soln of **25** (456 mg, 0.594 mmol) in 4:1 (v/v) CH₃CN-water (11 mL) was treated with 90% aq CF₃COOH (2.3 mL) warmed to 50 °C and stirred until the TLC analysis (EtOAc) showed the complete disappearance of the starting material (4 h). The mixture was concentrated under diminished pressure and repeatedly co-evaporated with toluene (5 × 20 mL). The residue was partitioned between brine (20 mL) and EtOAc (40 mL) and the aq phase extracted with EtOAc (3 × 40 mL). The organic phases were collected, dried (MgSO₄), concentrated under diminished pressure to give a residue (285 mg), that was directly subjected to a flash chromatographic purification, eluting with 4:6 hexane-EtOAc, to give **27** (228 mg, 86% yield) as a colourless syrup. NMR data were in agreement with those of the sample previously prepared by us.^{6b}

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Table 1: ¹H NMR parameters (δ, ppm; *J*, Hz) of the non-reducing unit of **9-11**, **14**, **16-20** and **22-25**

| Compound | Solvent | H-1' | H-2' | H-3' | H-4' | H-6'a | H-6'b | <i>J</i> _{1',2'} | <i>J</i> _{2',3'} | <i>J</i> _{3',4'} | <i>J</i> _{6'a,6'b} |
|-----------|--------------------|------|------|------|------|-------|-------|---------------------------|---------------------------|---------------------------|-----------------------------|
| 9 | CDCl ₃ | 4.86 | 3.61 | 3.89 | 4.11 | 3.59 | 3.50 | 7.9 | 9.4 | 3.7 | 10.4 |
| 10 | CD ₃ CN | 4.92 | 3.51 | 4.02 | 5.33 | 3.70 | 3.38 | 7.9 | 9.8 | 2.4 | 11.0 |
| 11 | CD ₃ CN | 5.15 | 3.91 | -- | 5.06 | 3.65 | 3.31 | 6.2 | -- | -- | 10.2 |
| 14 | CD ₃ CN | 4.86 | 3.38 | 3.60 | 3.79 | 3.72 | 3.58 | 8.0 | 9.1 | 9.1 | 10.1 |
| 16 | CDCl ₃ | 4.93 | 3.42 | 3.97 | 5.41 | 3.42 | 3.25 | 7.9 | 10.0 | 3.4 | 10.8 |
| 17 | CDCl ₃ | 5.04 | 4.04 | -- | 5.09 | 3.57 | 3.34 | 7.3 | -- | -- | 10.6 |
| 18 | CDCl ₃ | 5.24 | 3.45 | 3.45 | 5.10 | 3.33 | 3.27 | 8.2 | 3.3 | n.d. | 10.3 |
| 19 | CDCl ₃ | 5.31 | 3.63 | 5.24 | 3.42 | 3.52 | 3.46 | 8.2 | 3.6 | 3.3 | 10.6 |
| 20 | CDCl ₃ | 5.31 | 3.42 | 5.21 | 5.08 | 3.42 | 3.24 | 8.3 | 3.4 | 3.0 | 10.5 |
| 22 | CDCl ₃ | 4.86 | 3.52 | 4.00 | 3.87 | 3.53 | 3.45 | 7.9 | 9.9 | 3.4 | 10.3 |
| 23 | CDCl ₃ | 4.82 | 3.82 | 3.82 | 4.05 | 3.57 | 3.41 | 7.6 | n.d. | n.d. | 10.1 |
| 24 | CD ₃ CN | 4.91 | 4.31 | -- | 3.74 | 3.61 | 3.55 | 7.5 | -- | -- | 10.6 |
| 25 | CDCl ₃ | 5.17 | 3.51 | 3.99 | 3.73 | 3.55 | 3.42 | 8.4 | 3.4 | 3.2 | 10.2 |

Table 2: Selected ¹³C NMR signals (δ, ppm) of the disaccharide derivatives **9-11, 14** and **16-25**.

| Compound | Solvent | C-1 | C-2 | C-3 | C-4 | C-5 | C-6 | C-1' | C-2' | C-3' | C-4' | C-5' | C-6' |
|-----------|--------------------|-------|-------|-------|-------|-------|------|-------|-------|--------------------|-------|-------|------|
| 9 | CDCl ₃ | 105.5 | 74.4 | 77.8 | 75.0 | 77.2 | 65.5 | 98.9 | 78.8 | 78.6 | 67.2 | 100.3 | 64.7 |
| 10 | CD ₃ CN | 106.4 | 75.7 | 76.6 | 76.5 | 77.7 | 66.1 | 100.3 | 79.2 | 78.3 | 82.7 | 99.8 | 65.3 |
| 11 | CD ₃ CN | 106.6 | 76.0 | 78.8 | 76.5 | 77.9 | 66.2 | 99.6 | 76.5 | 155.3 | 99.4 | 99.5 | 70.3 |
| 14 | CD ₃ CN | 106.9 | 76.2 | 78.4 | 76.9 | 77.8 | 66.2 | 99.7 | 82.8* | 82.0 ^{a*} | 74.1 | 99.9 | 74.4 |
| 16 | CDCl ₃ | 105.5 | 74.5* | 77.8* | 76.4 | 77.6* | 64.8 | 98.4 | 74.3* | 69.6* | 68.8* | 99.5 | 64.1 |
| 17 | CDCl ₃ | 106.1 | 75.0* | 77.6 | 75.1* | 76.9 | 65.3 | 100.0 | 80.2 | 197.2 | 74.9* | 99.9 | 64.2 |
| 18 | CDCl ₃ | 106.2 | 75.5 | 77.7 | 75.2 | 77.3 | 65.4 | 96.4 | 74.3 | 69.3* | 69.2* | 101.6 | 64.9 |
| 19 | CDCl ₃ | 105.6 | 74.8* | 78.0 | 74.7* | 77.7 | 65.3 | 96.2 | 73.8* | 71.6 | 68.4 | 101.4 | 66.2 |
| 20 | CDCl ₃ | 105.8 | 74.8 | 77.9* | 75.1 | 77.8* | 65.2 | 95.9 | 73.1 | 68.5* | 68.4* | 100.5 | 65.2 |
| 21 | CDCl ₃ | 105.9 | 74.7 | 77.8 | 75.4 | 77.5 | 65.4 | 96.1 | 74.7 | 71.6 | 69.6 | 102.3 | 65.9 |
| 22 | CDCl ₃ | 105.4 | 74.3 | 77.8 | 74.6 | 77.3 | 65.2 | 99.1 | 79.6 | 70.7 | 77.5 | 100.8 | 64.7 |
| 23 | CDCl ₃ | 105.2 | 74.0 | 77.6 | 74.8 | 76.9 | 65.4 | 99.1 | 79.4 | 79.2 | 75.1 | 100.6 | 64.5 |
| 24 | CD ₃ CN | 106.5 | 75.8 | 78.4 | 77.4 | 77.7 | 66.2 | 100.9 | 82.0 | 202.3 | 82.0 | 101.6 | 64.9 |
| 25 | CDCl ₃ | 106.4 | 74.9 | 77.9 | 75.2 | 76.3 | 65.7 | 97.3 | 78.5* | 70.1 | 78.3* | 103.5 | 66.2 |

* Assignments may have to be interchanged